



Figure 1. Stereoscopic view of a computer-generated space-filling drawing, based on the X-ray coordinates of molecule 3, with an included benzene molecule. van der Waals radii of 1.2 Å for H, 1.85 Å for C, and 1.4 Å for O were used. The benzene is clamped at the edge of 3. Four other benzenes in lattice positions are not shown; two intrude into the cavity of 3, while the other two are outside the cage. The X-ray data were collected at -100 °C to avoid crystal decomposition, probably benzene loss. The crystal of 3 obtained was triclinic, PI, Z=2, a=15.1221 (29) Å, b=15.5931 (30) Å, c=15.7000 (41) Å, V=3463 (1) $Å^3$. A total of 5598 reflections were refined to a final R_w 0.141. Each unit cell contains one left-handed twist molecular complex as shown and one more right-handed mirror image of it.

in 66% yield from tri-p-anisylcarbinol.

The dimeric coupling reaction of 2 failed completely under standard and high-dilution coupling conditions (Cu(OAc)2 in pyridine^{1c} and Cu(I)-TMEDA-O₂⁶). However, we have devised a new copper coupling procedure that is extremely mild and efficient and that we have used for related couplings. Treatment of 2 (515 mg, 1.23 mmol) in O₂-free pyridine (1230 mL) with anhydrous CuCl (12.1 g, 123 mmol) and anhydrous CuCl₂ (2.0 g, 15 mmol) for 48 h at 0 °C gave the cage dimer 3, mp 180 °C dec, in 35% yield after isolation by preparative plate chromatography (2 mm silica, R_f 0.85, CH₂Cl₂ eluent) and crystallization from benzene. The ¹H NMR in CD₂Cl₂ showed an AA'BB' pattern for the aromatic hydrogens (7.10, 6.88 ppm, J = 8.5 Hz), while the propargyl methylenes and the bridgehead methyls were singlets at 4.78 and 2.05 ppm, respectively.

The crystals of 3 from benzene were suitable for an X-ray structure determination. The picture derived from the crystal structure (Figure 1) shows that one benzene molecule is tightly clamped in the cavity and shows no disorder and two others are more loosely held in the cavity, while two others occupy lattice positions around molecule 3. Furthermore, as Figure 1 shows, the molecule 3 is twisted by rotation of one triphenylethane unit relative to the other. We show the molecule with a left-handed helical twist, but the unit cell actually contains one left- and one right-handed molecule, with interlocking acetylene chains. The "dihedral" twist angles between phenyl rings across the C3 rotation axis^{1d} are 28°, 35°, and 41°. The lack of a crystallographic C_3 axis reflects the presence of the three unsymmetrically included benzenes. This twisting brings the two triphenylethane units together, clamping the tightly held benzene molecule in place. In molecular models the untwisted structure, with the triphenylethane units further apart, is more open and more easily admits the benzene molecule.

The further elaboration of this type of cage molecule must involve the attachment of catalytic and water solubilizing groups. It will also be of interest to see whether with bulkier guest molecules inside the cavity the twist angle is smaller, as expected. The highly efficient one-pot synthesis should make it possible to explore all these questions.

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Supplementary Material Available: Crystallographic data, ORTEP, atomic coordinates, bond lengths, bond angles, anisotropic temperature factors, hydrogen coordinates and temperature factors, and reflections, of the cage molecule (47 pp). Ordering information is given on any current masthead page.

A-(Modified B₆)-

B- $[\omega$ -amino(ethylamino)]- β -cyclodextrin as an Artificial B₆ Enzyme for Chiral Aminotransfer Reaction

Iwao Tabushi,* Yasuhisa Kuroda, Masahiko Yamada, and Hideyuki Higashimura

> Department of Synthetic Chemistry Kyoto University, Sakyo-ku, Kyoto, 606 Japan

Ronald Breslow

Department of Chemistry, Columbia University New York, New York 10027 Received April 17, 1985

In Kyoto we have studied the regiospecific introduction of functional groups into cyclodextrin for the construction of enzyme mimics;1 at Columbia pyridoxamine has been attached to cyclodextrin to promote substrate binding, and flexible chains carrying amino groups have been attached to pyridoxamine to promote proton transfers, including stereospecific amino acid synthesis.² We now wish to report the coalescence of these research lines: the synthesis of B₆-dependent aminotransferase model 3 (see Chart I): (1) converting certain keto acids into amino acids under mild conditions in water, (2) consisting of two cooperating units, a modified B₆ coenzyme grouping and an ω-amino apoenzyme grouping, (3) exhibiting almost exclusive L-chiral induction from keto-prochiral groupings (see eq 1).

Thus, A,B-capped cyclodextrin 1 was converted to the B₆ model 3 via diiodide 23 by using pyridoxamine thiol^{2a,b} under the con-

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Chart I. Preparation of Regioisomeric Artificial B_{ϵ} Enzymes from A,B-Capped Cyclodextrin

Table I. Characteristic Spectra of 3a

electronic, nm (ϵ) (aq, pH 8.0) 250 (ϵ 4400), 324 (6600) 400-MHz ¹H NMR; (D₂O, pH 4.0) SC₆H δ 2.55 dd (J = 12.6, 7.6 Hz), 2.82 dd (J = 12.6, 1.7 Hz). 3b: 2.64 dd (J = 11.2 Hz, 7.7), and 3.01 dd (J = 11.2 Hz, 1.0). 100-MHz ¹³C NMR; (D₂O, pH 4.0) 157.7, 144.4, 137.5, 132.5, 103.6, 102.8, 84.5, 83.2, 82.9, 74.9, 74.6, 74.3, 74.1, 73.9, 73.7, 73.5, 69.3, 62.5. 62.3, 50.2, 46.7, 37.3, 36.9, 33.7, 33.0, 16.9

Table II. Chiral Induction in Amino Acids Formed via the Artificial B₆ Catalysis^a

R-COCO₂H	L/D	
$R = PhCH_2$	98/2	
CH ₂	95/5 ^b	
Ph	98/2 ^b	

^aAt 30 °C, 2 M K₂HPO₄-KH₂PO₄ buffer, pH 8.0. ^b Recorded ratio is lowest limit.

ditions listed in Chart I. The crude mixture free from ethylenediamine was applied on a CM-25 Sephadex column. From $0.12-0.15~M~NH_4HCO_3$ gradient via the usual workup was obtained 64 mg of 3·HCl (15% yield).

Separation of the two regioisomers 3a and 3b was successfully carried out again by CM-25 column chromatography of a prepurified 3a + 3b mixture (60/40 ratio) by the use of a long, thin column (5 × 1500 mm) eluted with aqueous NH₄HCO₃ (0-0.1 M gradient). The observed elution diagram obtained by measuring intensities of the absorption at 324 nm (B_6 chromophore) of each 3-mL fraction clearly shows that the two regioisomers are satisfactorily separated. The latter eluent was recrystallized from aqueous EtOH twice, giving pure A,B regioisomer 3a, while from the earlier fractions, pure B,A regioisomer 3b, was similarly obtained. Both compounds showed pyridoxamine characteristic CH₂ signals and ethylenediamine CH₂ signals in expected intensities, but the signals of 3a SCH₂ at C₆ were found at 2.82 and 2.55 ppm while those of 3b were at 3.01 and 2.64 ppm. Less than 1% of the contaminating signal could be seen (see further Table I).

The aminotransfer reaction smoothly proceeded between 3a and keto acids 4a—c under the conditions described in eq 1, giving the corresponding amino acids 5a—c, respectively. The amino acids formed were further converted to the 3,5-dinitrobenzamides 6a—c which were acidified (pH 3) and extracted with ether (10 mL \times 3). The condensed ether layer (150 μ L) was analyzed by HPLC by using a Sumipax OA-1000 chiral column eluted with a MeOH/H₂O/CH₃CO₂H mixture (95/5/1 vol). The two peaks of each racemic amino acid derivative were clearly resolved. Under

the conditions, total yields of amino acids were ca. $40\%^4$ and the enantiomeric excess observed for L-amino acid was very high, as listed in Table II.

The observed high chiral induction for 3a may be understood on the basis of stereospecific participation^{2e} of the ω -NH₂ grouping toward the prochiral azomethine plane fixed nearly perpendicular toward the N-lone pair during the prototropy (see eq 3), due to

the host-guest multiple recognition.^{1,5} The mechanism is further supported by the acceleration of prototropy caused by the assisting NH₂ group on the B ring. The observed prototropy rate ratio followed spectroscopically, k (3) to k (free B₆), was ca. 2000–2200 in the L-phenylalanine formation reaction, in a good agreement with the observed chiral induction.⁶

(4) The present yield of 6 is neither corrected for the conversion in acylation (which may not be exactly the same for D- and L-amino acid) nor the extraction efficiency.

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(6) Or, rate ratio k (3) to k (B₆ cyclodextrin) was 10–11. Detailed analysis of complex multiphase kinetics will appear in a full-length article.

Formation of CO₂ and a Four-Membered 1,3-Dimetallacycle by Deoxygenation of a Ketone with [Rh(CO)₂Cl]₂

J. William Suggs,* Michael J. Wovkulich, and Ken S. Lee

Department of Chemistry, Brown University Providence, Rhode Island 02912

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8-substituted quinoline ligands have proven useful in studies of carbon-hydrogen and carbon-carbon bond activation by transition metals.¹ They allow specific ligand bonds to be brought into a metal's coordination sphere whereupon reaction can occur. We now wish to describe an unexpected example of deoxygenation of a ketone by a rhodium carbonyl complex in an 8-substituted quinoline ligand.

When 8-quinolinyl phenyl ketone (1) and [Rh(CO)₂Cl]₂ (2) are mixed in benzene at 25 °C under argon (mole ratio 1:1) the initially yellow solution darkens to a deep red color within 1 h.² At this time CO₂ evolution begins, as shown by a new IR band in the solution at 2330 cm⁻¹ and by trapping of the evolved gas using Ca(OH)₂. The CO₂ evolution continues for approximately 24 h. Absorption of the CO₂ on Ascarite indicates slightly less than 1 mol of CO₂ is formed per mole of ketone. Deep red crystals of a new compound 3, slowly formed and after 3 days these were collected (67% yield).³ They can be handled in air, but slowly

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⁽²⁾ Upon mixing, a new species with IR bands at 2098, 2080, 2026, 2011, and 1652 cm⁻¹ forms. The decrease in the ketone band from 1668 cm⁻¹ in 1 to 1652 cm⁻¹ is consistent with bidentate N,O coordination of 1 to a rhodium species.

^{(3) 3:} mp 172–174 °C dec; IR (CH₂Cl₂) 2079 (s), 2057 (m), 2011 (s) cm⁻¹; NMR (250 MHz, ¹H, CD₂Cl₂) & 8.28 (dd, 1 H), 7.86 (dd, 1 H), 7.50 (dd, 1 H), 7.40 (dd, 1 H), 7.4-7.3 (m 6 H), 7.01 (dd, 1 H).